



## REVIEW ARTICLE

# Umbilical cord mesenchymal stem cells and their secretome: a new frontier in orthopedic medicine

Tito Sumarwoto<sup>1,2\*</sup> , Romaniyanto<sup>1,3</sup> , Mujaddid Idulhaq<sup>1,4</sup> , Asep Santoso<sup>1,5</sup>   
and Sholahuddin Rhatomy<sup>6</sup> 

<sup>1</sup>Department of Orthopaedic and Traumatology, Soeharso Orthopaedic Hospital, Sebelas Maret University Faculty of Medicine, Surakarta, Indonesia

<sup>2</sup>Division of Upper Extremity, Hand, and Microsurgery, Soeharso Orthopaedic Hospital, Sebelas Maret University Faculty of Medicine, Surakarta, Indonesia

<sup>3</sup>Division of Spine, Soeharso Orthopaedic Hospital, Surakarta, Indonesia

<sup>4</sup>Division of Musculoskeletal Tumor, Soeharso Orthopaedic Hospital, Surakarta, Indonesia

<sup>5</sup>Division of Adult Reconstruction, Soeharso Orthopaedic Hospital, Surakarta, Indonesia

<sup>6</sup>Division of Adult Reconstruction, Dr. Soeradji Tirtonegoro General Hospital, Klaten – Faculty of Medicine, Universitas Gadjah Mada, Yogyakarta, Indonesia

### \* Correspondence Author:

 [tito.sumarwoto@rso.go.id](mailto:tito.sumarwoto@rso.go.id)

Cite this article as: Sumarwoto T, Romaniyanto, Idulhaq M, Santoso A, Rhatomy S. Umbilical cord mesenchymal stem cells and their secretome: a new frontier in orthopedic medicine. Univ Med 2026;45:135-152

Date of first submission, November 5, 2025

Date of final revised submission, April 7, 2026

Date of acceptance, April 22, 2026

## ABSTRACT

Umbilical cord mesenchymal stem cells (UC-MSCs) have gained significant attention in regenerative medicine due to their unique biological properties, including high proliferation capacity, low immunogenicity, and potent immunomodulatory effects. These characteristics make UC-MSCs particularly promising for orthopedic applications, where the repair and regeneration of musculoskeletal tissues such as bone, cartilage, tendons, ligaments, and nerves are critical for restoring function. The secretome of UC-MSCs—comprising bioactive molecules such as exosomes, cytokines, and growth factors—offers a powerful, cell-free therapeutic option through paracrine signaling, further enhancing their therapeutic potential. A literature search was conducted in major databases (PubMed, ScienceDirect, SpringerLink, Google Scholar) for English articles from 2010–2025 using keywords related to UC-MSCs and orthopedic regeneration. This review explores the role of UC-MSCs and their secretome in orthopedic tissue repair, focusing on their application in bone healing, cartilage regeneration, tendon-ligament repair, and nerve regeneration with their innovative delivery. Despite the promising potential of UC-MSC therapies, several challenges remain, including regulatory hurdles, long-term safety concerns, and the scalability of cell-based and secretome-based therapies for widespread clinical use. Although umbilical cord MSCs are not yet widely applied in clinical practice, increasing evidence suggests that they offer significant therapeutic potential, especially in the treatment of autoimmune and neurodegenerative diseases. The UC-MSCs and their secretome represent a transformative approach in orthopedics, offering new avenues for treating complex musculoskeletal injuries and degenerative diseases. Ongoing advancements in this field will likely unlock their full potential, making them viable options for clinical use in the near future.

**Keywords:** Umbilical cord-mesenchymal stem cells (UC-MSCs), secretome, orthopedics, neurodegenerative diseases

### Abbreviations

ACL: Anterior Cruciate Ligament

BDNF: Brain-Derived Neurotrophic Factor  
BMPs: Bone Morphogenetic Proteins  
ECM: Extracellular Matrix  
EMA: European Medicines Agency  
EVs: Extracellular Vesicles  
FDA: Food and Drug Administration  
GDNF: Glial Cell-Derived Neurotrophic Factor  
GMP: Good Manufacturing Practices  
HSCs: Hematopoietic Stem Cells  
HUVECs: Human Umbilical Vein Endothelial Cells  
IGF: Insulin-Like Growth Factor  
IL-10: Interleukin-10  
IL-6: Interleukin-6  
MHC: Major Histocompatibility Complex  
miRNA: MicroRNA  
MSCs: Mesenchymal Stem Cells  
NGF: Nerve Growth Factor  
NK cells: Natural Killer Cells  
OA: Osteoarthritis  
PDGF: Platelet-Derived Growth Factor  
RNA: Ribonucleic Acid  
TGF- $\beta$ : Transforming Growth Factor-Beta  
TNF- $\alpha$ : Tumor Necrosis Factor-Alpha  
UC-MSCs: Umbilical Cord-Derived Mesenchymal Stem Cells  
VEGF: Vascular Endothelial Growth Factor

## INTRODUCTION

Orthopedic medicine, that may be applied for addressing musculoskeletal disorders, such as bone fractures, osteoarthritis, tendon injuries, intervertebral disc degeneration, and peripheral nerve lesions, has currently modality therapeutic options on surgical intervention, mechanical support, and pharmacological management. These therapeutic options may not always lead to optimal healing and can be associated with complications such as infection, implant rejection, or limited long-term efficacy, and often result in chronic pain, disability, and decreased quality of life.<sup>(1)</sup> As a result, there is a growing need for advanced, biologically based therapeutic approaches that can enhance tissue regeneration and repair.<sup>(2,3)</sup>

In recent years, mesenchymal stem cells (MSCs), particularly umbilical cord-mesenchymal stem cells (UC-MSCs), have emerged as a promising option for addressing orthopedic challenges. Their ability to differentiate into bone, cartilage, and tendon cells and Schwann cell-like cells makes them particularly suitable for treating a wide range of musculoskeletal injuries and conditions.<sup>(4)</sup> Furthermore, UC-MSCs secrete bioactive molecules (the secretome) that enhance tissue

healing by modulating inflammation, promoting angiogenesis, and stimulating cell proliferation and differentiation.<sup>(5)</sup>

Umbilical cord mesenchymal stem cells and their secretome are being explored in preclinical and clinical studies for their potential to accelerate bone healing, regenerate cartilage, repair damaged tendons and ligaments, as well as to repair and regenerate peripheral nerves. Their non-invasive harvesting process, robust regenerative capacity, and low risk of immune rejection make them particularly promising in the field of orthopedics, where effective and long-lasting tissue repair is crucial for patient recovery.<sup>(6,7)</sup>

The aim of this review was to provide a comprehensive analysis of the use of MSCs, specifically UC-MSCs, with their secretome, as a potential for regenerative medicine in treatment approaches for orthopedic tissue repair. This paper highlights the unique properties of UC-MSCs that make them suitable for regenerative therapy and innovative delivery, and also discusses the challenges and limitations of UC-MSC therapies in orthopedics.

## METHODS

This narrative review synthesized current evidence on the therapeutic use of UC-MSCs and

their secretome in orthopedic applications, including bone, cartilage, tendon, joint, and nerve repair. A comprehensive search of PubMed, Scopus, ScienceDirect, and Google Scholar was performed for articles published between January 2015 and March 2025 using the terms: “umbilical cord mesenchymal stem cells” OR “UC-MSCs” AND “secretome” AND (“orthopedics” OR “bone” OR “cartilage” OR “tendon” OR “joint” OR “nerve” OR “musculoskeletal”) AND (“regeneration” OR “repair” OR “healing”).

Eligible articles focused on UC-MSCs or their secretome in orthopedics-related tissues or conditions, were original studies, reviews, or clinical trials in English, and reported mechanistic or therapeutic outcomes. Studies on non-orthopedic systems, abstracts, editorials, conference posters, and unpublished material were excluded. The following data were extracted: study type and model (in vitro, in vivo, clinical), source and characterization of UC-MSCs, application site (e.g., bone defect, osteoarthritis, nerve lesion), role of secretome or exosomes, outcome measures (histological, molecular, functional), key findings and limitations. The data were summarized narratively, categorized by

tissue type (bone, cartilage, tendon, nerve), and discussed in the context of regenerative mechanisms and translational potential. The study selection process for a systematic review of umbilical cord-derived mesenchymal stem cells (UC-MSCs) in orthopedics was conducted according to the PRISMA guidelines, as diagrammed in Figure 1. Records were identified through database searching of PubMed, Scopus, ScienceDirect, and Google Scholar between 2015 and 2025 ( $n = 280$ ). After removal of duplicate records ( $n = 50$ ), 230 reports were screened for retrieval. Thirty reports could not be retrieved due to access limitations or unavailability of full-text articles, leaving 200 reports for full-text eligibility assessment based on predefined inclusion and exclusion criteria. Of these, 96 reports were excluded (31 non-orthopedic studies, 25 abstracts without full text, 20 editorials or conference posters, and 20 articles on irrelevant topics). A total of 104 studies met the eligibility criteria and were included in the qualitative synthesis, comprising studies on bone regeneration, cartilage repair, tendon/ligament regeneration, and nerve regeneration (Fig. 1).

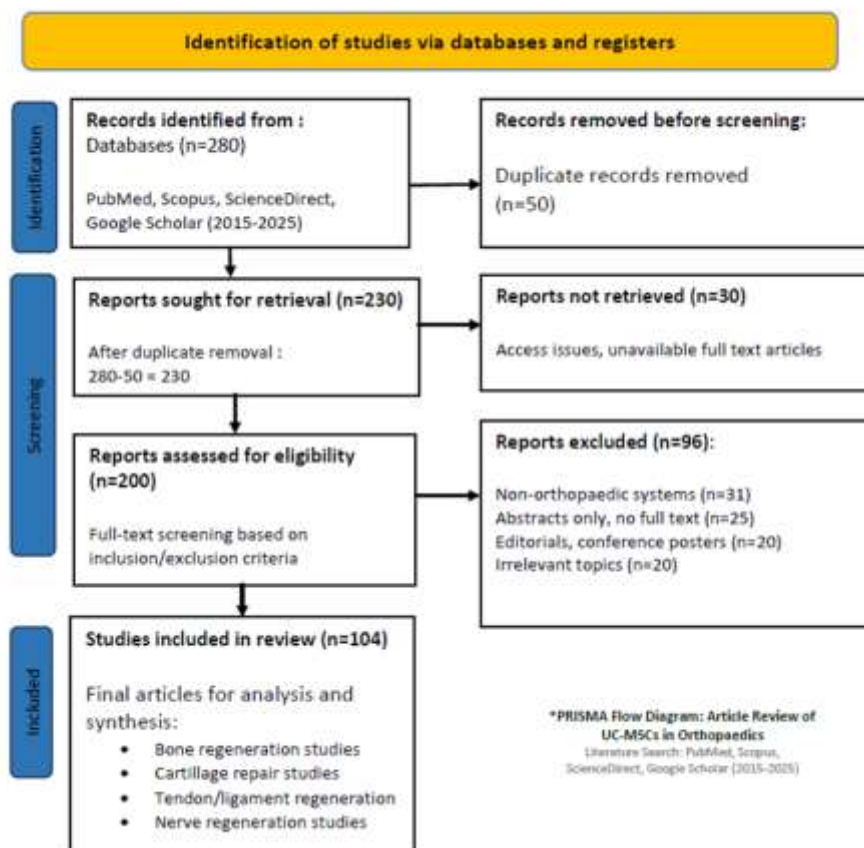


Figure 1. PRISMA flow diagram: article review of UC-MSCs in orthopedics

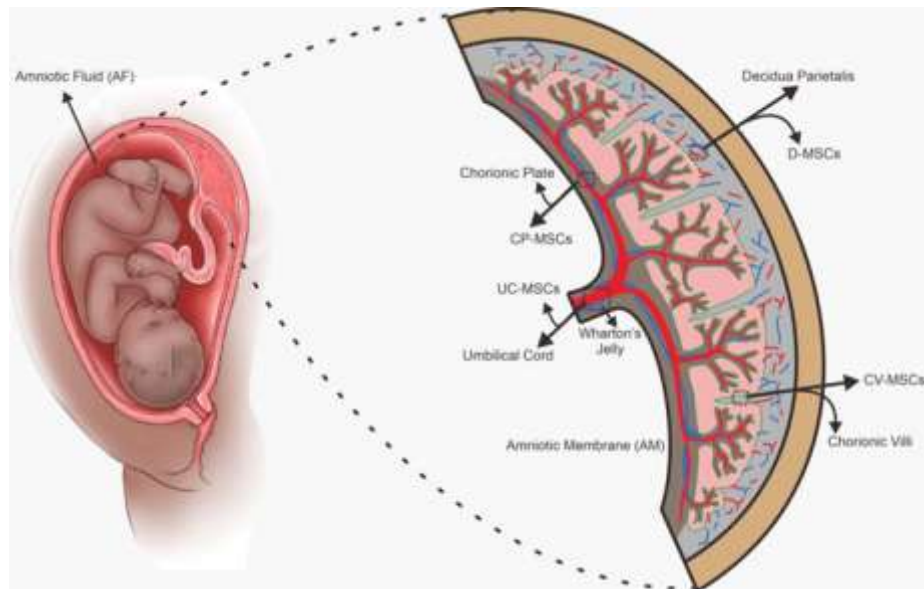


Figure 2. Anatomical structure of the fetus in the uterus and a cross section of the placenta showing the various sources of mesenchymal stem cells (MSCs). The placenta, along with associated tissues such as the umbilical cord and amniotic membrane, is an important source of stem cells that can be used for regenerative therapies.

The different types of stem cells shown in the image include : UC-MSCs (mesenchymal stem cells from the umbilical cord), CP-MSCs (stem cells from the chorionic plate), CV-MSCs (stem cells from the chorionic villi), D-MSCs (stem cells from the parietal decidua/maternal layer), AF-MSCs (stem cells from amniotic fluid), and stem cells from Wharton's Jelly a mucoïd tissue in the umbilical cord that is rich in MSCs. The extraembryonic tissues such as the placenta not only play a role in supporting fetal growth but are also a potential source of multipotent stem cells for future medical applications

### Characteristics of UC-MSCs

The umbilical cord is a highly accessible, ethically favorable source of mesenchymal stem cells (MSCs), as it is routinely discarded after birth and can be collected non-invasively without risk to mother or infant. UC-MSCs are mainly isolated from Wharton's jelly, a gelatinous matrix within the cord that is particularly rich in MSCs. As illustrated in Figure 2, the placenta and related extraembryonic tissues also provide diverse MSC populations, including UC-MSCs from Wharton's jelly, CP-MSCs from the chorionic plate, CV-MSCs from chorionic villi, D-MSCs from the maternal decidua, and AF-MSCs from amniotic fluid, all of which are routinely discarded material that may be transformed into valuable, ethically acceptable reservoirs of multipotent stem cells for regenerative medicine.<sup>(8)</sup>

Isolation of UC-MSCs starts with cord collection after delivery, followed by cleaning, disinfection, and dissection to obtain Wharton's jelly. MSCs are then isolated either by enzymatic digestion (e.g., collagenase) to release cells from the matrix or by explant culture, where tissue pieces are placed in medium and the cells migrate out. The harvested cells are subsequently expanded in vitro in appropriate culture media to

obtain sufficient numbers for therapeutic applications.<sup>(9,10)</sup>

Umbilical cord mesenchymal stem cells (UC-MSCs) have potent immunomodulatory capacity, suppressing proliferation and activation of T cells, B cells, and NK cells to reduce inflammation. They also secrete anti-inflammatory cytokines, creating a favorable healing microenvironment and enabling their use in allogeneic transplantation with minimal need for intensive immunosuppression.<sup>(11)</sup>

Umbilical cord mesenchymal stem cells have the capacity to differentiate into various mesenchymal lineages (Fig.2), including osteocytes (bone cells), chondrocytes (cartilage cells), tenocytes (tendon cells),<sup>(12)</sup> and Schwann-cell like cells.<sup>(13)</sup> This makes them highly suitable for orthopedic applications, as they can contribute directly to the repair of musculoskeletal tissues. Their osteogenic differentiation supports bone healing and regeneration, while their chondrogenic potential is crucial for cartilage repair in conditions such as osteoarthritis. Their ability to differentiate into tenocytes and also into Schwann-cell like cells allows UC-MSCs to play a role in tendon and ligament repair, as well as in peripheral nerve repair and regeneration.<sup>(14,15)</sup>

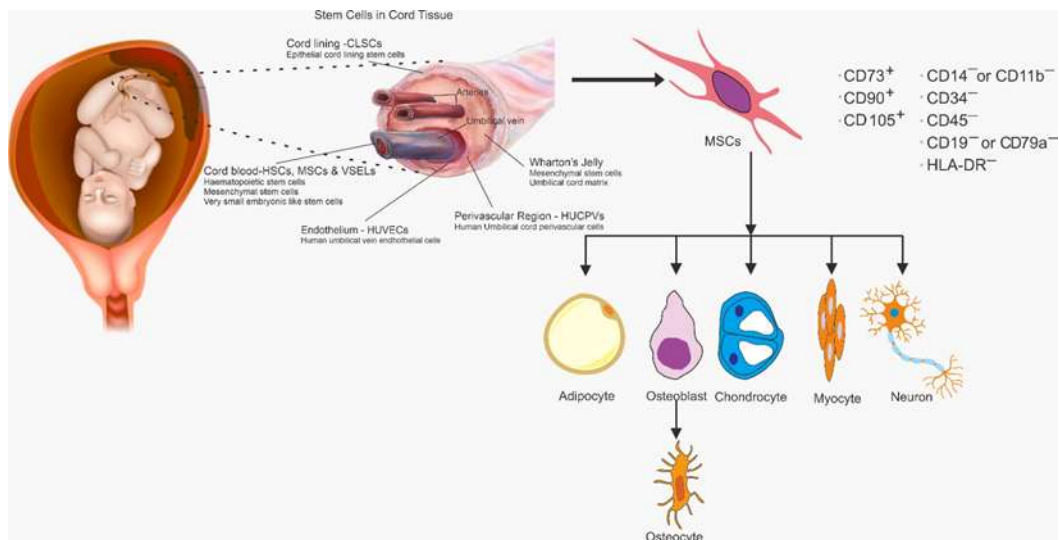


Figure 3. This image shows the different types of stem cells that can be obtained from human umbilical cord tissue and their differentiation capabilities. The umbilical cord, previously considered medical waste, is now known to be a rich source of mesenchymal stem cells (MSCs) and other cell types with high therapeutic potential. The main sites that produce stem cells include: Cord lining/LSCs (contains epithelial stem cells from the inner lining of the umbilical cord), Cord blood (contains hematopoietic stem cells/HSCs, mesenchymal stem cells/MSCs, and very small embryonic-like stem cells/VSELs), Wharton's Jelly (contains MSCs in the mucoid matrix of the umbilical cord), Perivascular region/HUCPVs (contains stem cells from the area surrounding blood vessels), Endothelium/HUVECs (Endothelial cells from the human umbilical vein). Immunophenotypic features of MSCs shown include: positive for CD73+, CD90+, CD105+; negative for CD14-/CD11b-, CD34-, CD45-, CD19-/CD79a-, HLA-DR-. These MSCs are able to differentiate into various cell types including: adipocytes/fat cells, osteoblasts and osteocytes/bone cells, chondrocytes/cartilage cells, myoblasts/myocytes (muscle cells), neurons/nerve cells. With their multipotent differentiation capabilities and favorable immunological profile, these cells have great potential for applications in regenerative therapies and cell-based medicine

Compared with other MSC sources, UC-MSCs show lower immunogenicity, largely due to reduced expression of MHC class I and II molecules. This makes them particularly suitable for allogeneic, off-the-shelf therapies with less need for strict donor–recipient matching.<sup>(16)</sup> Umbilical cord mesenchymal stem cells have a higher proliferation rate compared to MSCs derived from sources such as bone marrow or adipose tissue. They can be expanded more rapidly in culture, allowing for the generation of larger cell numbers in a shorter time frame (Fig.3). This is particularly advantageous for clinical applications that require high doses of MSCs for effective treatment.<sup>(17)</sup> Additionally, UC-MSCs maintain their stemness and differentiation potential for a longer period compared to other MSC sources, making them more viable for long-term regenerative therapies.<sup>(18)</sup>

Umbilical cord mesenchymal stem cells secrete a variety of bioactive molecules, including growth factors and cytokines, which play a crucial role in promoting tissue repair and regeneration.<sup>(5,19)</sup> These secreted factors (UC-

MSC secretome), include vascular endothelial growth factor (VEGF), transforming growth factor-beta (TGF- $\beta$ ), and various interleukins, which help stimulate angiogenesis, reduce inflammation, and enhance the proliferation and differentiation of resident cells in damaged tissues.<sup>(20)</sup> The UC-MSC secretome is particularly important for paracrine signaling, where UC-MSCs influence nearby cells and tissues to promote healing, even in situations where direct differentiation into target cells is not the primary mode of action.<sup>(21,22)</sup>

Overall, the combination of low immunogenicity, high proliferation rates, and a rich secretome makes UC-MSCs a highly attractive candidate for use in regenerative medicine, particularly in the field of orthopedics, where effective tissue repair and regeneration are paramount.<sup>(23,24)</sup>

### The role of UC-MSCs in orthopedic applications

Umbilical cord mesenchymal stem cells (UC-MSCs) contribute significantly to

osteogenesis by differentiating into osteoblasts that produce bone matrix and support mineralization (Fig. 4). They secrete growth factors such as BMPs, VEGF, and TGF- $\beta$ , which promote osteogenic differentiation, stimulate angiogenesis, and recruit additional regenerative cells to the injury site, thereby enhancing bone remodeling and repair.<sup>(25)</sup>

The UC-MSCs have shown great promise in treating bone fractures, particularly in cases of delayed union or non-union where normal healing processes are impaired.<sup>(7,26)</sup> Umbilical cord mesenchymal stem cells can be applied directly to the fracture site or delivered through scaffolds, where they assist in forming new bone tissue by differentiating into osteoblasts and releasing their regenerative secretome.<sup>(27)</sup> Studies have shown that UC-MSCs accelerate bone healing by promoting the deposition of new bone matrix, reducing inflammation, and stimulating angiogenesis, which improves the blood supply to

the injured area.<sup>(28,29)</sup> In preclinical and clinical settings, UC-MSCs have demonstrated the potential to enhance healing in cases of severe fractures or in patients with compromised healing capacity, such as those with osteoporosis or advanced age.<sup>(26,30)</sup>

Cartilage repair is a significant challenge in orthopedics, particularly for degenerative diseases such as osteoarthritis, where the cartilage in the joints breaks down and leads to pain and loss of function.<sup>(31,32)</sup> The UC-MSCs have been explored as potential therapy for regenerating damaged cartilage due to their ability to differentiate into chondrocytes and secrete bioactive molecules (Fig.5) that promote tissue repair.<sup>(33,34)</sup> When applied to areas of cartilage damage, UC-MSCs can differentiate into chondrocytes and contribute to the regeneration of the extracellular matrix, which is essential for cartilage function and durability.<sup>(35)</sup>

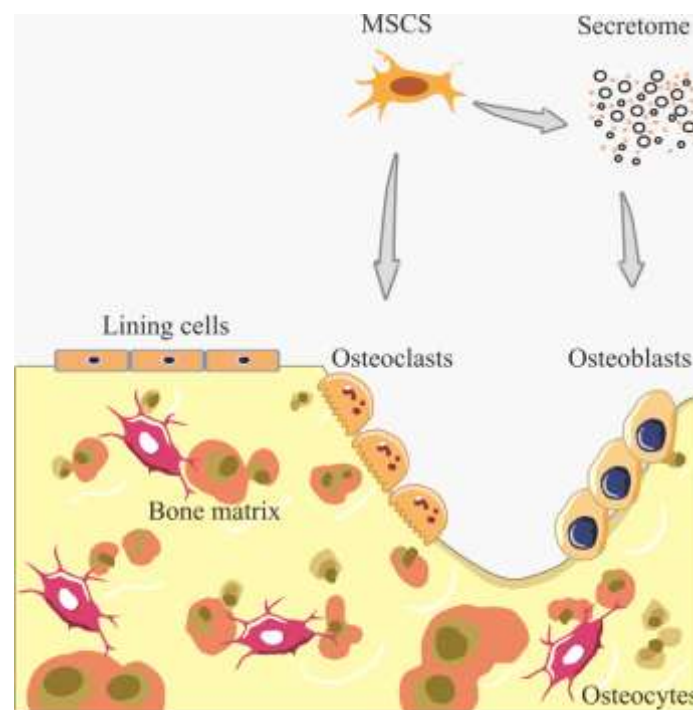


Figure. 4 This illustration provides a visual representation of the role of Mesenchymal Stem Cells (MSCs) and their secretome in the process of bone remodeling. MSCs are shown as the origin of both direct cellular influence and the production of a biologically active substance (secretome), play crucial roles in intercellular communication and regulation within the bone microenvironment.

MSCs and their secretome impact osteoclasts that are responsible for bone resorption, breaking down the bone matrix and facilitating the removal of old or damaged bone tissue; and osteoblasts, involved in bone formation, synthesizing new bone matrix and contributing to bone regeneration. Over time, some osteoblasts become embedded within the matrix they produce, differentiating into osteocytes, which act as mechanosensors and regulators of bone remodeling. Lining cells on the bone surface originate from osteoblasts and regulate mineral exchange, as well as support bone remodeling activities. Within the bone matrix, osteocytes and various other cells maintain homeostasis and coordinate the dynamic balance between bone resorption and formation

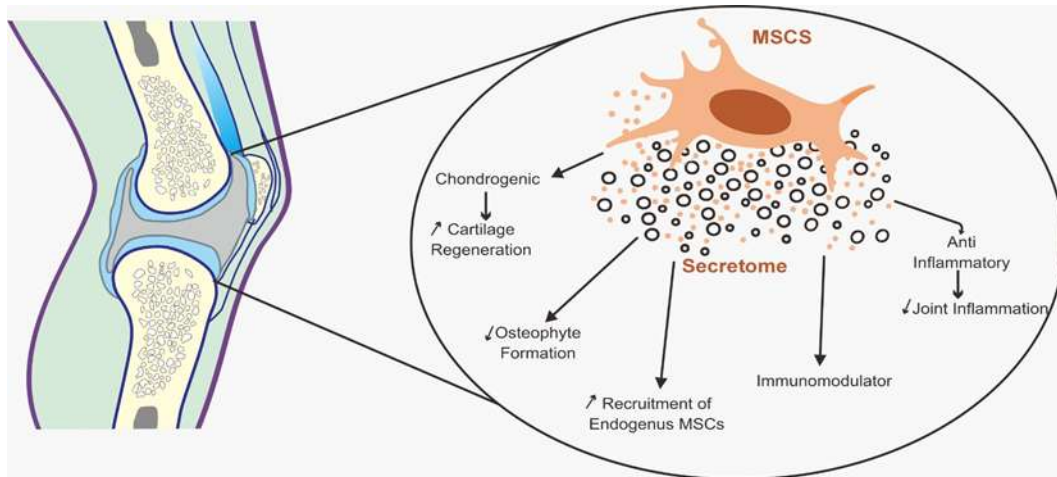


Figure 5. This illustration highlights the therapeutic potential of Mesenchymal Stem Cells (MSCs) and their secretome in the context of joint disease, particularly osteoarthritis (OA).

The left image shows a degenerative joint—likely the knee—where cartilage breakdown and inflammation are key pathological features. On the right, the focus shifts to the MSCs and their secretome, which encompasses a variety of bioactive molecules including cytokines, growth factors, and extracellular vesicles.

**Chondrogenic Activity:** The secretome promotes chondrogenesis, leading to enhanced cartilage regeneration, a critical factor in restoring joint function and preventing further degeneration.

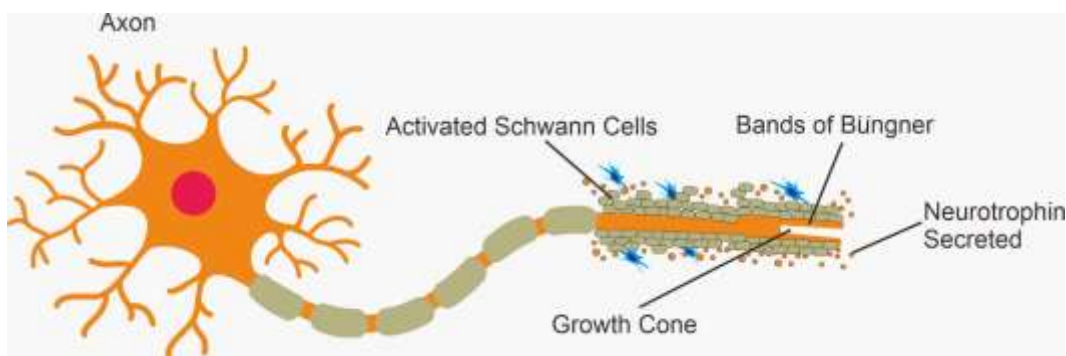
**Reduction in Osteophyte Formation:** By modulating the joint environment, MSC secretome helps suppress the formation of osteophytes (bony outgrowths), which are a hallmark of OA and contribute to joint stiffness and pain.

**Recruitment of Endogenous MSCs:** The factors released by MSCs can attract the body's own stem cells to the site of injury, amplifying regenerative processes and aiding long-term healing.

**Immunomodulation:** The secretome exhibits immunomodulatory effects, helping to balance the immune response and prevent chronic inflammation that would otherwise damage joint tissues.

**Anti-Inflammatory Effects:** Through the secretion of anti-inflammatory mediators, MSC secretome reduces joint inflammation, providing symptomatic relief and slowing disease progression.

This diagram underscores how MSC-derived secretome acts as a cell-free therapy capable of addressing multiple pathological aspects of joint degeneration—structural damage, inflammation, and impaired repair—making it a promising strategy for regenerative medicine in osteoarthritis and related joint disorders.



**Figure 6.** This illustration depicts the critical steps and cellular components involved in **peripheral nerve regeneration** following injury, highlighting the role of Schwann cells and neurotrophic signaling in axonal repair. After injury, Schwann cells become activated and begin to proliferate and clear debris from the damaged axon and play a pivotal role in guiding axonal regrowth. **Bands of Büngner** are longitudinal columns formed by aligned Schwann cells and their basal lamina, provide a **physical and molecular scaffold** that directs the regenerating axon toward its target, ensuring proper orientation and connectivity. At the tip of the regenerating axon, the growth cone explores the microenvironment, responding to molecular cues for directional growth.

**Neurotrophin secretion**, the secretion from activated Schwann cells (NGF, BDNF, and GDNF) enhance neuronal survival, stimulate axon elongation, and support synaptic reconnection.

The UC-MSCs undergo chondrogenic differentiation in response to specific signals such as TGF- $\beta$  and insulin-like growth factor (IGF). These factors promote the expression of key chondrogenic markers, including collagen type II and aggrecan, which are essential components of the cartilage matrix.<sup>(36,37)</sup> Additionally, UC-MSCs' paracrine activity—mediated through the secretion of anti-inflammatory cytokines and growth factors—can modulate the local environment to reduce cartilage degradation and stimulate tissue repair.<sup>(24,38)</sup> Preclinical studies and clinical trials have demonstrated that UC-MSCs can improve cartilage repair, reduce pain, and improve joint function in osteoarthritis patients, suggesting their potential as a novel therapeutic approach for cartilage regeneration.<sup>(39)</sup>

Tendon and ligament injuries, such as rotator cuff tears and anterior cruciate ligament (ACL) ruptures, are common orthopedic conditions that often require surgical intervention.<sup>(40)</sup> Umbilical cord mesenchymal stem cells have been explored as biological therapy to improve tendon and ligament repair due to their ability to differentiate into tenocytes and their potential to enhance the healing process.<sup>(41,42)</sup> When applied to injured tendons or ligaments, UC-MSCs can promote tissue regeneration by increasing collagen synthesis, reducing inflammation, and stimulating the repair of the extracellular matrix. Their ability to secrete growth factors, such as platelet-derived growth factor (PDGF) and TGF- $\beta$ , helps in creating a favorable environment for tendon and ligament healing.<sup>(43)</sup>

In preclinical studies, UC-MSCs have shown success in improving the healing of tendon and ligament injuries. Animal models with induced tendon or ligament injuries that were treated with UC-MSCs demonstrated improved biomechanical properties, reduced inflammation, and enhanced tissue regeneration compared to untreated controls.<sup>(44)</sup> Early-phase clinical trials have also shown promising results, with UC-MSC-treated patients experiencing faster recovery times, improved functional outcomes, and reduced re-injury rates. These findings suggest that UC-MSCs could become an important adjunct to surgical repair techniques for tendon and ligament injuries.<sup>(45)</sup>

Spinal disc degeneration and the need for spinal fusion surgery represent significant challenges in orthopedic care. Current treatments for degenerative disc disease often involve spinal

fusion, which eliminates motion between vertebrae but does not address the underlying degeneration of the intervertebral discs.<sup>(46)</sup> Umbilical cord mesenchymal stem cells offer a potential regenerative solution by promoting the regeneration of the nucleus pulposus and annulus fibrosus. The UC-MSCs can differentiate into discogenic cells and produce extracellular matrix proteins, such as collagen and proteoglycans, that are essential for disc function.<sup>(47,48)</sup>

In the context of spinal fusion, UC-MSCs can enhance bone healing and fusion at the surgical site. By promoting osteogenesis and releasing bioactive molecules that stimulate bone formation, UC-MSCs may help achieve a more robust and successful fusion.<sup>(49)</sup> Early studies suggest that UC-MSCs could reduce recovery times and improve outcomes in spinal fusion surgeries, potentially offering a regenerative approach to disc degeneration and improving the long-term function of the spine.<sup>(50,51)</sup>

Wallerian degeneration is a process that occurs after peripheral nerve injury in the peripheral nerve repair and regeneration, where the part of the axon distal to the injury site degenerates and the myelin sheath breaks down. This process is essential for clearing damaged cells and preparing the nerve for regeneration, but it also involves significant inflammation and immune cell activation.<sup>(52,53)</sup> The UC-MSCs play a critical role in modulating this process by exerting immunomodulatory and anti-inflammatory effects that can enhance the repair of peripheral nerves.<sup>(45,53)</sup>

Umbilical cord mesenchymal stem cells have been shown to influence the Wallerian degeneration process by reducing the inflammatory response and promoting an environment conducive to nerve regeneration (Fig.6). Through the secretion of bioactive molecules such as interleukin-10 (IL-10) and TGF- $\beta$ , UC-MSCs can attenuate the activation of immune cells such as macrophages, which are involved in clearing debris but can also contribute to further nerve damage if overly activated.<sup>(20)</sup> Additionally, UC-MSCs release neurotrophic factors, such as brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF), which support the survival and growth of neurons during the regeneration process.<sup>(54)</sup> By modulating the local immune response and promoting neuroprotection, UC-MSCs help to create an environment that facilitates axonal regrowth,

remyelination, and ultimately functional recovery of the nerve after injury.<sup>(55,56)</sup>

Peripheral nerve injuries and lesions, such as those resulting from trauma, compression, or surgical procedures, often lead to a loss of motor or sensory function, presenting a significant clinical challenge.<sup>(57)</sup> Conventional treatments for peripheral nerve injuries, including nerve grafts or surgical repairs, often have limited success, especially in cases of large nerve gaps or delayed treatment. UC-MSCs offer a promising therapeutic alternative due to their regenerative properties and ability to promote nerve repair.<sup>(45,58)</sup>

The UC-MSCs can enhance peripheral nerve regeneration by promoting the differentiation of Schwann cells, which are critical for axonal regrowth and remyelination. Schwann cells provide physical support to growing axons and secrete factors that aid in nerve repair. When UC-MSCs are applied to injured peripheral nerves, they can promote Schwann cell proliferation and function, thus supporting the natural regenerative processes of the nerve.<sup>(54,59)</sup>

Additionally, UC-MSCs can secrete neuroprotective and angiogenic factors, such as VEGF, which help to restore blood supply to the injured area, further enhancing the repair process. This is especially important in cases of nerve crush injuries or other forms of ischemic damage, where restoring proper blood flow is essential for nerve survival and regeneration.<sup>(60)</sup>

In cases of more severe nerve lesions, such as nerve transection or extensive damage, UC-MSCs can be applied in combination with bioengineered scaffolds or nerve conduits to bridge the nerve gap and facilitate axonal regeneration. The scaffolds provide physical guidance for the regenerating axons, while UC-MSCs contribute to the biochemical signals necessary for nerve regrowth. This combination has shown promise in preclinical models, with evidence of improved nerve regeneration and functional recovery compared to standard surgical repairs alone.<sup>(61,62)</sup>

The UC-MSCs have also been shown to reduce scar tissue formation at the injury site, which is a common barrier to successful nerve regeneration. By limiting the extent of fibrosis and promoting a pro-regenerative environment, UC-MSCs can enhance the likelihood of successful nerve repair and functional recovery.<sup>(63,64)</sup>

In preclinical studies and animal models of peripheral nerve injury, UC-MSCs have demonstrated the ability to accelerate nerve regeneration, improve axonal regrowth, and

enhance functional recovery.<sup>(65)</sup> Early clinical trials are exploring the use of UC-MSCs in treating peripheral nerve injuries, particularly in cases where conventional treatments have failed or are insufficient. The results thus far suggest that UC-MSCs hold great potential for improving outcomes in peripheral nerve repair, offering a new avenue of treatment for patients with traumatic nerve injuries or chronic nerve lesions.<sup>(45,66)</sup>

### UC-MSCs secretome: clinical studies for the treatment of orthopedic conditions

The **secretome** refers to the collection of bioactive molecules secreted by cells, including proteins, lipids, nucleic acids, and other factors that influence surrounding cells and tissues.<sup>(28)</sup> In the context of UC-MSCs, the secretome is composed of a variety of substances that contribute to tissue repair and regeneration through paracrine signaling mechanisms.<sup>(67,68)</sup>

The major components of the UC-MSC secretome include exosomes, cytokines, and growth factors. Exosomes are small extracellular vesicles that carry proteins, RNA, and microRNAs (miRNAs). These vesicles play a key role in cell communication and deliver signals to target cells to modulate their behavior.<sup>(69)</sup> Cytokines are signaling proteins that regulate immune responses and inflammation, such as IL-10, IL-6, and tumor necrosis factor-alpha (TNF- $\alpha$ ).<sup>(70)</sup> Growth factors are molecules such as VEGF, TGF- $\beta$ , PDGF, and IGF, that stimulate tissue repair, angiogenesis, and cell proliferation.<sup>(71)</sup> These components collectively contribute to the therapeutic potential of the UC-MSC secretome in promoting healing in musculoskeletal injuries and diseases.

The UC-MSC secretome exerts its regenerative effects primarily through paracrine signaling, in which secreted factors from UC-MSCs influence neighboring cells and tissues to promote repair and regeneration without the need for direct cell-to-cell contact.<sup>(72)</sup> The key mechanisms of action of the secretome include anti-inflammatory effects, anti-fibrotic effects, and angiogenic effects. The secretome contains cytokines and growth factors, such as IL-10 and TGF- $\beta$ , that help reduce inflammation by modulating immune responses (anti-inflammatory effects). These factors inhibit the activation of pro-inflammatory immune cells, such as macrophages and T-cells, which is critical in preventing chronic inflammation and creating a conducive environment for tissue repair.<sup>(28,73)</sup>

The UC-MSC secretome can reduce scar tissue formation (anti-fibrotic effects) by limiting the deposition of extracellular matrix proteins, such as collagen, that contribute to fibrosis. This is particularly important in the context of musculoskeletal injuries, where excessive scar tissue can impede functional recovery.<sup>(23,74)</sup>

The secretome is rich in angiogenic factors, such as VEGF and PDGF, which promote the formation of new blood vessels (angiogenic effects). This enhanced vascularization is crucial for delivering oxygen and nutrients to damaged tissues, thus accelerating the healing process in bone, cartilage, and soft tissue injuries.<sup>(75)</sup>

### **Secretome in bone and cartilage repair**

Studies have shown that the UC-MSC secretome plays a pivotal role in bone and cartilage regeneration.<sup>(27)</sup> Preclinical research has demonstrated that the secretome can enhance osteogenesis and chondrogenesis through several pathways.<sup>(27,35)</sup> The UC-MSC secretome promotes bone healing by stimulating the proliferation and differentiation of osteoblasts, the cells responsible for bone formation. It also enhances the recruitment of endogenous stem cells to the site of injury and promotes the release of growth factors that accelerate bone repair. Studies in animal models have shown that the application of UC-MSC-derived exosomes can improve bone healing in fractures, especially in cases of delayed or impaired healing.<sup>(28,68)</sup>

The UC-MSC secretome has demonstrated the ability to stimulate chondrocyte activity and promote the deposition of cartilage matrix, which is essential for cartilage repair. In models of osteoarthritis, the secretome has been shown to reduce inflammation, slow cartilage degradation, and improve the regeneration of cartilage tissue. This suggests a potential therapeutic application in treating cartilage defects and degenerative joint diseases.<sup>(76,77)</sup> The secretome in peripheral nerve regeneration and repair. The UC-MSCs secretome plays a crucial role in peripheral nerve repair and regeneration following injury or lesion. Instead of relying solely on the differentiation of MSCs into nerve cells, researchers have found that their paracrine effects—specifically the bioactive factors that they secrete—are key drivers of regeneration.<sup>(66)</sup>

The crucial roles in peripheral nerve repair and regeneration, including neuroprotection, neuroregeneration, immunomodulation, angiogenesis, and recruitment - activation of

endogenous cells. The MSCs secrete factors such as brain-derived neurotrophic factor (BDNF), nerve growth factor (NGF), and glial cell-derived neurotrophic factor (GDNF). These support neuron survival, prevent apoptosis, and maintain axonal integrity after injury.<sup>(78)</sup> Exosomes and EVs from MSCs can transfer miRNAs that modulate gene expression in injured neurons and Schwann cells. These promote axon sprouting, axon elongation, and myelin sheath restoration.<sup>(79)</sup> The MSC secretome reduces local inflammation by secreting anti-inflammatory cytokines (e.g., IL-10, TGF- $\beta$ ) and suppressing pro-inflammatory cytokines (e.g., TNF- $\alpha$ , IL-1 $\beta$ ). This creates a more favorable environment for nerve regeneration and minimizes secondary damage.<sup>(11,80)</sup> The UC-MSCs release VEGF and other angiogenic factors that promote the formation of new blood vessels. Improved vascularization supports nutrient and oxygen supply to regenerating nerve tissue.<sup>(81)</sup> The UC-MSCs secretome can recruit Schwann cells, macrophages, and endogenous stem cells to the injury site. Schwann cells are essential for guiding axonal regrowth and remyelination.<sup>(82)</sup>

### **Advantages over whole-cell therapy**

The UC-MSC secretome offers several practical advantages over whole-cell therapy, making it an attractive alternative for regenerative medicine applications. The secretome can be easily stored, handled, and administered without the need for complex cell culture techniques. Unlike live cells, the secretome can be stored as an "off-the-shelf" product in frozen or lyophilized (freeze-dried) form, allowing for greater convenience and broader clinical applicability.<sup>(83)</sup>

While UC-MSCs themselves exhibit low immunogenicity, the secretome has an even lower risk of triggering an immune response because it lacks the cellular components (e.g., cell membranes, nuclei) that could be recognized as foreign by the recipient's immune system. This makes the secretome an ideal candidate for allogeneic applications (using donor-derived products) without the need for immunosuppression.<sup>(5,73)</sup>

The secretome avoids several potential risks associated with cell-based therapies. Live stem cells have a theoretical risk of uncontrolled proliferation and tumor formation (tumorigenicity) when transplanted into patients. The secretome, consisting of non-cellular components, eliminates this risk.<sup>(84)</sup> The

secretome can be derived from discarded umbilical cords, making it ethically uncontroversial, unlike some other sources of stem cells.<sup>(23)</sup> The secretome is easy to regulate and standardize for therapeutic use, providing a more controlled and predictable treatment approach compared to cell-based therapies, where the behavior of live cells can vary depending on patient conditions.<sup>(85)</sup>

### **Orthopedic medicine: innovative stem cells and secretome delivery**

In regenerative orthopedics, injectable hydrogels serve as protective carriers for therapeutic agents, shielding them from rapid enzymatic degradation and clearance *in vivo*. They enable controlled and sustained release of bioactive components over days to weeks, improving therapeutic efficacy while reducing dosing frequency. Hydrogels can also promote integration of the therapeutic payload into host tissue by facilitating cell migration, matrix deposition, and vascular ingrowth at the defect site.<sup>(86)</sup> Table 1 briefly overviews the clinical use of the UC-MSC secretome in orthopedics, mainly intra-articular injections for knee osteoarthritis, including product type, study design, patient number, dosing, and key outcomes, and shows that these treatments appear safe, and reduce pain and improve function, while bone-defect and disc-degeneration applications of UC-MSC secretome/exosomes remain preclinical (Table 1). Some injectable hydrogels are three-dimensional, water-rich polymeric networks that closely mimic the structural and biochemical properties of the extracellular matrix (ECM). Their high water content and tunable mechanical properties create a biocompatible and hydrated environment, making them ideal for encapsulating living cells, stem cell secretome, or bioactive molecules. By resembling the native ECM, hydrogels provide physical support and biochemical cues that enhance cell survival, proliferation, and differentiation in regenerative applications.<sup>(91)</sup>

Formulations such as chitosan-based gels, collagen solutions, and Pluronic F127 undergo a sol-gel transition at physiological temperature (thermosensitive hydrogels). These can be injected as liquids into irregular cartilage defects, where they solidify *in situ*, enabling precise, localized delivery of stem cells or secretome.<sup>(92)</sup>

Hydrogels integrated with nanoparticles (e.g., hydroxyapatite, silica, or polymeric nanocarriers) allow dual delivery of cells and

growth factors (nanocomposite hydrogels). This strategy has shown promise in enhancing bone healing by providing both osteoconductive scaffolding and sustained biochemical stimulation.<sup>(93)</sup>

Injectable hydrogels offer a minimally invasive delivery method that conforms to complex tissue geometries without requiring open surgery. Once injected, they undergo gelation at the target site, ensuring localized retention of therapeutic agents and reducing systemic side effects. Their tunable degradation rates and release kinetics allow for customized treatment regimens tailored to specific orthopedic conditions.<sup>(86)</sup>

The scaffold-based localized delivery involves the use of biodegradable, three-dimensional matrices that function both as structural frameworks for tissue regeneration and as reservoirs for therapeutic agents such as stem cell secretome.<sup>(94)</sup> These scaffolds mimic the architecture of native extracellular matrix (ECM), providing an environment that supports cell adhesion, proliferation, and differentiation while gradually degrading to be replaced by newly formed tissue.<sup>(95)</sup>

In orthopedic applications, scaffolds serve a dual role for mechanical stability, in that they firstly fill and reinforce bone or cartilage defects, maintain the space needed for tissue ingrowth, and prevent collapse of the defect site, and secondly they provide biological stimulation when loaded with UC-MSCs or their secretome scaffolds act as a localized delivery depot and gradually release regenerative signals (e.g., growth factors, cytokines, exosomes) that promote angiogenesis, osteogenesis, or chondrogenesis.<sup>(96)</sup>

Collagen and fibrin scaffolds (natural polymer scaffolds), valued for their biocompatibility and bioactivity, can be impregnated with UC-MSCs or secretome for bone regeneration, enabling both osteoconductive support and paracrine signaling.<sup>(97)</sup> Bio-ceramic scaffolds that are composed of hydroxyapatite,  $\beta$ -tricalcium phosphate, or other calcium phosphate ceramics, are osteoconductive and mechanically strong, making them suitable for load-bearing bone defect repair.<sup>(98)</sup> Scaffolds can be fabricated with patient-specific shapes and controlled porosity to match irregular defect geometries (3D-printed porous scaffolds) by using additive manufacturing for ensuring a snug fit and optimal tissue integration.<sup>(99)</sup> Scaffold-based delivery offers the synergistic benefit of combining

mechanical reinforcement with targeted biological stimulation. This integration enhances structural integrity, guides tissue regeneration, and supports long-term healing without the need for additional fixation materials once the scaffold degrades.<sup>(100)</sup>

### **Challenges and limitations of UC-MSC therapies in orthopedics**

One of the primary challenges facing the clinical application of UC-MSC therapies is navigating the complex regulatory landscape and addressing ethical concerns. Regulatory challenges arise in the standardization of UC-MSC-based products for clinical use. The process of isolating, expanding, and delivering UC-MSCs is complex, and ensuring consistent quality, potency, and purity of cells across different batches is difficult. Regulatory agencies, such as the US FDA and European Medicines Agency (EMA), have established strict guidelines for cell-based therapies, including ensuring that manufacturing processes are compliant with Good Manufacturing Practices (GMP). This includes defining stringent protocols for cell source, handling, storage, and expansion, but these vary between countries, making global standardization a challenge.<sup>(101)</sup> The secretome presents additional regulatory hurdles. As a cell-free product composed of secreted factors, the secretome lacks standardization in terms of defining its components, dosage, and storage. Regulatory agencies must also address how to regulate these biologically complex products to ensure safety and efficacy.<sup>(28,64)</sup> Although UC-MSCs are derived from non-invasive sources, such as discarded umbilical cords after childbirth, there are still ethical concerns to consider. These primarily revolve around ensuring informed consent from the donor's parents and ensuring that the donation process is voluntary and free from exploitation.<sup>(102)</sup>

While UC-MSCs are considered ethically more acceptable than other stem cell sources (such as embryonic stem cells), public perception and concerns over the potential commercialization of human biological materials remain. There may also be questions surrounding the long-term risks of using stem cell-derived products in clinical applications, which may require further ethical review.<sup>(84)</sup>

Though UC-MSC therapies have shown promise in early clinical trials, there remain several safety concerns that need to be addressed, particularly regarding long-term efficacy. While UC-MSCs are known for their low

immunogenicity, there remains a small risk of triggering an immune response in recipients, particularly in allogeneic (donor-derived) applications. This is why UC-MSC therapies are being closely monitored for any signs of immune rejection or inflammatory reactions. However, due to the immunomodulatory nature of UC-MSCs, these risks are typically low and manageable.<sup>(28)</sup>

There is a theoretical risk of tumor formation when using live stem cells in regenerative medicine, as stem cells have the ability to proliferate and differentiate. Although UC-MSCs are not considered highly tumorigenic, their proliferative capacity still raises concerns, especially in long-term follow-up studies. Researchers have focused on minimizing this risk by selecting and culturing cells in a way that avoids uncontrolled growth or mutations, but vigilance is required in clinical applications.<sup>(84)</sup>

While short-term results from UC-MSC therapies in orthopedics are promising, there is still limited data on the long-term outcomes of these treatments. It remains uncertain whether the beneficial effects, such as improved tissue regeneration or pain relief, persist over time, or if additional treatments are required to maintain these effects. Long-term studies are critical to understanding the durability of UC-MSC-based therapies.<sup>(26,39)</sup>

Another significant challenge for UC-MSC-based therapies is the ability to scale up production for widespread clinical use while maintaining quality and consistency. Expanding UC-MSCs for large-scale clinical applications presents several challenges. UC-MSCs need to be cultured and expanded under strict GMP conditions to ensure that the cells retain their therapeutic properties, but scaling this process can lead to changes in cell characteristics, such as reduced differentiation potential or altered secretory profiles. As demand for stem cell-based therapies grows, it will be necessary to develop reliable and efficient methods for expanding cells without compromising their quality. The production of the secretome requires careful control of the cell culture conditions to ensure that the correct therapeutic molecules are produced in sufficient quantities. This is complicated by the fact that the secretome's composition can vary depending on how UC-MSCs are cultured, how long they are cultured, and the environmental conditions they are exposed to.<sup>(23)</sup>

Ensuring quality control during the production process is critical to the success of UC-

MSC-based therapies. Variability in the biological properties of UC-MSCs, such as their immunomodulatory or differentiation potential, can occur due to differences in donor characteristics (e.g., age, health) or in vitro culture conditions. This variability can make it difficult to produce a consistent, standardized product that performs the same way across different batches and patients.<sup>(103)</sup> For secretome-based therapies, ensuring the reproducibility of the composition of secreted factors (such as exosomes, cytokines, and growth factors) is an ongoing challenge. The complexity of the secretome and its dynamic nature during cell culture makes it difficult to characterize and standardize for clinical use.<sup>(64)</sup> Moreover, methods for quality control, such as potency assays, need to be developed and validated to ensure that the secretome maintains its therapeutic efficacy across different production runs.<sup>(104)</sup>

## CONCLUSION

Umbilical cord mesenchymal stem cells are a promising tool in orthopedics, offering non-invasive sourcing, low immunogenicity, and high proliferation. They show strong potential in bone, cartilage, tendon, ligament, and nerve regeneration, aided by their immunomodulatory, anti-inflammatory, and angiogenic effects. The UC-MSC-derived secretome, rich in bioactive molecules, provides a cell-free alternative with easier storage and lower tumorigenic risk. While early studies are encouraging, future research must address regulatory, safety, and manufacturing challenges to ensure clinical translation.

## Acknowledgment

The authors would like to acknowledge the Department of Orthopaedics and Traumatology, Soeharso Orthopaedics Hospital, Surakarta, Indonesia, for supporting the publication.

## Authors' Contributions

TTO suggested the original manuscript's topic and wrote its first draft. ROM and MJD made revisions and edits to the manuscript. RTM and ASP provided contextual input on the updated paper. TTO oversaw the manuscript and made significant edits to the draft. Every author has approved the manuscript's final edit.

## Conflict of Interest

The authors declare no conflict of interest.

## Funding

The authors received no funding for this study.

## Data Availability Statement

This is a narrative review article. No new data were generated or analyzed during the preparation of this manuscript.

## Declaration of AI Usage in Scientific Writing

The authors declare that they have not used AI-generated work in this manuscript.

## REFERENCES

1. Das S, Thakur A, Datta A, Sahoo A, Bandyopadhyay S, Sah AK. Advances in regenerative medicine for orthopedic injuries: a comprehensive review. *Cureus* 2025;17:e79860. doi: 10.7759/cureus.79860.
2. Bulut D, Sharabidze Z. Regenerative medicine in orthopaedic surgery: pioneering advances and their applications. *EMJ Innov* 2025;9:82-94. doi: 10.33.590/emjinnov/FGDS3814
3. Liang W, Zhou C, Bai J, et al. Current advancements in therapeutic approaches in orthopedic surgery: a review of recent trends. *Front Bioeng Biotechnol* 2024;12:1328997. doi: 10.3389/fbioe.2024.1328997.
4. Trapana J, Weinerman J, Lee D, et al. Cell-based therapy in the treatment of musculoskeletal diseases. *Stem Cells Transl Med* 2024;13:959-78. doi: 10.1093/stcltm/szae049.
5. Drobiova H, Sindhu S, Ahmad R, Haddad D, Al-Mulla F, Al Madhoun A. Wharton's jelly mesenchymal stem cells: a concise review of their secretome and prospective clinical applications. *Front Cell Dev Biol* 2023;11:1211217. doi: 10.3389/fcell.2023.1211217.
6. Szablowska-Gadomska I, Rudziński S, Dymowska M. Secretome of mesenchymal stromal cells as a possible innovative therapeutic tool in facial nerve injury treatment. *Biomed Res Int* 2023;2023:8427200. doi: 10.1155/2023/8427200.
7. Smolinska V, Csobonyeiova M, Zamborsky R, Danisovic L. Stem cells and their derivatives: an implication for the regeneration of nonunion fractures. *Cell Transplant* 2023;32:9636897231183530. doi: 10.1177/09636897231183530
8. Advani D, Barragan JV, Statache G, Kadri N, Kohli N. Upcycled mesenchymal stem cells: repurposing biological waste towards sustainable.

- Cell Engineering Connect 2025;1:1–14. doi : 10.69709/CellEngC.2025.101060.
9. Xu M, Xu J, Cheng D, et al. Isolation of umbilical cord-derived mesenchymal stem cells with high yields and low damage. *J Vis Exp* 2024;209:e66835. doi: 10.3791/66835.
  10. Kestendjieva S, Chervenkov M, Oreshkova T, Mourdjeva M, Stoyanova E. Mesenchymal stromal/stem cells isolated by explant culture method from Wharton's jelly and subamion possess similar biological characteristics. *Appl. Sci* 2024;14:8036. doi: 10.3390/app14178036.
  11. Vohra M, Arora SK. Mesenchymal stem cells—the master immunomodulators. *Explor Immunol* 2023;3:104–22. doi: 10.37349/ei.2023.00092.
  12. Rehman A, Nigam A, Laino L, et al. Mesenchymal stem cells in soft tissue regenerative medicine: a comprehensive review. *Medicina (Lithuania)* 2023;59:1–20. doi: 10.3390/medicina59081449.
  13. Choi SJ, Park SY, Shin YH, et al. Mesenchymal stem cells derived from Wharton's jelly can differentiate into Schwann cell-like cells and promote peripheral nerve regeneration in acellular nerve grafts. *Tissue Eng Regen Med* 2021;18:467–78. doi: 10.1007/s13770-020-00329-6.
  14. Xu Q, Hou W, Zhao B, et al. Mesenchymal stem cells lineage and their role in disease development. *Mol Med* 2024;30:207. doi: 10.1186/s10020-024-00967-9.
  15. Wei C, Guo Y, Ci Z, Li M, Zhang Y, Zhou Y. Advances of Schwann cells in peripheral nerve regeneration: from mechanism to cell therapy. *Biomed Pharmacother* 2024;175:116645. doi :10.1016/j.biopha.2024.116645.
  16. Cequier A, Vázquez FJ, Vitoria A, et al. The systemic cellular immune response against allogeneic mesenchymal stem cells is influenced by inflammation, differentiation and MHC compatibility: in vivo study in the horse. *Front Vet Sci* 2024;11:1391872. doi: 10.3389/fvets.2024.1391872.
  17. Hori A, Takahashi A, Miharu Y, et al. Superior migration ability of umbilical cord-derived mesenchymal stromal cells (MSCs) toward activated lymphocytes in comparison with those of bone marrow and adipose-derived MSCs. *Front Cell Dev Biol* 2024;12:1–13. doi: 10.3389/fcell.2024.1329218.
  18. Tian L, Wang W, Li X, et al. Whole transcriptome scanning and validation of negatively related genes in UC-MSCs. *Heliyon* 2024;10:e27996. doi: 10.1016/j.heliyon.2024.e27996.
  19. Li P. Comparative breakthrough: Umbilical cord mesenchymal stem cells vs bone marrow mesenchymal stem cells in heart failure treatment. *World J Cardiol* 2024;16:776–80. doi: 10.4330/wjc.v16.i12.776.
  20. García-Guerrero CA, Fuentes P, Araya MJ, et al. How to enhance mscs therapeutic properties? an insight on potentiation methods. *Stem Cell Res Ther* 2024;15:331. doi: 10.1186/s13287-024-03935-6.
  21. Seok J, Park H, Cetin E, Ghasroldasht MM, Liakath FB, Al-Hendy A. The potent paracrine effect of umbilical cord mesenchymal stem cells mediates mitochondrial quality control to restore chemotherapy-induced damage in ovarian granulosa cells. *Biomed Pharmacother* 2024;172:116263. doi: 10.1016/j.biopha.2024.116263.
  22. Shan Y, Zhang M, Tao E, et al. Pharmacokinetic characteristics of mesenchymal stem cells in translational challenges. *Signal Transduct Target Ther* 2024;9:1–27. doi: 10.1038/s41392-024-01936-8.
  23. Prado-Yupanqui JW, Ramírez-Orrego L, Cortez D, et al. The hidden power of the secretome: therapeutic potential on wound healing and cell-free regenerative medicine—a systematic review. *Int J Mol Sci* 2025;26:1–20. doi: 10.3390/ijms26051926.
  24. Zhidu S, Ying T, Rui J, Chao Z. Translational potential of mesenchymal stem cells in regenerative therapies for human diseases: challenges and opportunities. *Stem Cell Res Ther* 2024;15:266. doi : 10.1186/s13287-024-03885-z.
  25. Wang L, Ruan M, Bu Q, Zhao C. Signaling pathways driving MSC osteogenesis: mechanisms, regulation, and translational applications. *Int J Mol Sci* 2025;26:1311. doi: 10.3390/ijms26031311.
  26. Cui C, Lin F, Xia L, Zhang X. Mesenchymal stem cells therapy for the treatment of non-union fractures: a systematic review and meta-analysis. *BMC Musculoskelet Disord* 2025;26:245. doi: 10.1186/s12891-025-08365-w.
  27. Kangari P, Talaei-Khozani T, Razeghian-Jahromi I, Razmkhah M. Mesenchymal stem cells: amazing remedies for bone and cartilage defects. *Stem Cell Res Ther* 2020;11:1–21. doi: 10.1186/s13287-020-02001-1.
  28. Trigo CM, Rodrigues JS, Camões SP, Solá S, Miranda JP. Mesenchymal stem cell secretome for regenerative medicine: where do we stand? *J Adv Res* 2024;70:103–24. doi: 10.1016/j.jare.2024.05.004.
  29. Bian D, Wu Y, Song G, Azizi R, Zamani A. The application of mesenchymal stromal cells (MSCs) and their derivative exosome in skin wound healing: a comprehensive review. *Stem Cell Res Ther* 2022;13:1–17. doi: 10.1186/s13287-021-02697-9.
  30. Zhang Y, Fan M, Zhang Y. Revolutionizing bone defect healing: the power of mesenchymal stem cells as seeds. *Front Bioeng Biotechnol* 2024;12:1–18. doi: 10.3389/fbioe.2024.1421674.

31. Householder NA, Raghuram A, Agyare K, Thipapay S, Zumwalt M. A review of recent innovations in cartilage regeneration strategies for the treatment of primary osteoarthritis of the knee: intra-articular injections. *Orthop J Sports Med* 2023;11:1–20. doi: 10.1177/23259671231155950.
32. Liu Y, Shah KM, Luo J. Strategies for articular cartilage repair and regeneration. *Front Bioeng Biotechnol* 2021;9:1–10. doi: 10.3389/fbioe.2021.770655.
33. Liao ZK. Clinical research progress of umbilical cord blood mesenchymal stem cells in knee articular cartilage repair: a review. *Medicine (United States)* 2025;104:e41402. doi: 10.1097/MD.0000000000041402.
34. Piñeiro-Ramil M, Gómez-Seoane I, Rodríguez-Cendal AI, Fuentes-Boquete I, Díaz-Prado S. Mesenchymal stromal cells-derived extracellular vesicles in cartilage regeneration: potential and limitations. *Stem Cell Res Ther* 2025;16:11. doi: 10.1186/s13287-025-04135-6.
35. Wu KC, Chang YH, Ding DC, Lin SZ. Mesenchymal stromal cells for aging cartilage regeneration: a review. *Int J Mol Sci* 2024;25:12911. doi: 10.3390/ijms252312911.
36. Cho GH, Bae HC, Lee YJ, et al. Insulin-like growth factor 2 secreted from mesenchymal stem cells with high glutathione levels alleviates osteoarthritis via paracrine rejuvenation of senescent chondrocytes. *Biomater Res* 2025;29:1–16. doi: 10.34133/bmr.0152.
37. Anatolitou A, Sideri K, Mavrogenis A, et al. Cartilage extracellular matrix collagen type II and aggrecan expressions in rabbit cartilage following mesenchymal stem cell implantation. *German J Vet Res* 2024;4:197–207. doi: 10.51585/gjvr.2024.3.0110.
38. Peshkova M, Korneev A, Suleimanov S, et al. MSCs' conditioned media cytokine and growth factor profiles and their impact on macrophage polarization. *Stem Cell Res Ther* 2023;14:1–16. doi: 10.1186/s13287-023-03381-w.
39. Chen Y, Cheng RJ, Wu Y, Huang D, Li Y, Liu Y. Advances in stem cell-based therapies in the treatment of osteoarthritis. *Int J Mol Sci* 2024;25:394. doi: 10.3390/ijms25010394.
40. Leong NL, Kator JL, Clemens TL, James A, Enamoto-Iwamoto M, Jiang J. Tendon and ligament healing and current approaches to tendon and ligament regeneration. *J Orthop Res* 2020;38:7–12. doi: 10.1002/jor.24475.
41. Shen Y, Wang Y, Xu Y, et al. Therapeutic potential and mechanisms of umbilical cord mesenchymal stem cells differentiating into tendon cells and promotion of rotator cuff tendon-bone healing. *J Tissue Eng* 2025;16:20417314251315185. doi: 10.1177/20417314251315185.
42. Augustin G, Jeong JH, Kim MK, Hur SS, Lee JH, Hwang Y. Stem cell-based therapies and tissue engineering innovations for tendinopathy: a comprehensive review of current strategies and future directions. *Adv Ther (Weinh)* 2024;7:1–23. doi: 10.1002/adtp.202300425.
43. Wu J, Wu J, Liu Z, et al. Mesenchymal stem cell-derived extracellular vesicles in joint diseases: therapeutic effects and underlying mechanisms. *J Orthop Translat* 2024;48:53–69. doi: 10.1016/j.jot.2024.07.005.
44. Yang J, Liu Y, Wang M, et al. Repair effect of umbilical cord mesenchymal stem cells embedded in hydrogel on mouse insulinoma 6 cells injured by streptozotocin. *Polymers (Basel)* 2024;16:1845. doi: 10.3390/polym16131845.
45. Song S, Li C, Xiao Y, Ye Z, Rong M, Zeng J. Beyond conventional therapies: mscs in the battle against nerve injury. *Regen Ther* 2025;28:280–91. doi: 10.1016/j.reth.2024.12.017.
46. Natarajan RN, Andersson GBJ. Lumbar disc degeneration is an equally important risk factor as lumbar fusion for causing adjacent segment disc disease. *J Orthop Res* 2017;35:123–30. doi: 10.1002/jor.23283.
47. Huang H, Liu X, Wang J, et al. Umbilical cord mesenchymal stem cells for regenerative treatment of intervertebral disc degeneration. *Front Cell Dev Biol* 2023;11:1–11. doi: 10.3389/fcell.2023.1215698.
48. Ohnishi T, Homan K, Fukushima A, Ukeba D, Iwasaki N, Sudo H. A review: methodologies to promote the differentiation of mesenchymal stem cells for the regeneration of intervertebral disc cells following intervertebral disc degeneration. *Cells* 2023;12:2161. doi: 10.3390/cells12172161.
49. Kim Y hoon, Kim K won, Rhyu K won, et al. Bone fusion materials : past , present , and future. *Asian Spine J* 2025;1–10. doi: 10.31616/asj.2024.0520.
50. Keshavarz S, Alavi CE, Aghayan H, Jafari-Shakib R, Vojoudi E. Advancements in degenerative disc disease treatment: a regenerative medicine approach. *Stem Cell Rev Rep* 2025;21:1252–82. doi: 10.1007/s12015-025-10882-z.
51. Munda M, Velnar T. Stem cell therapy for degenerative disc disease: Bridging the gap between preclinical promise and clinical potential. *Biomol Biomed* 2024;24:210–8. doi: 10.17305/bb.2023.9518.
52. Tian R, Zhou Y, Ren Y, Zhang Y, Tang W. Wallerian degeneration: from mechanism to disease to imaging. *Heliyon* 2025;11:e40729. doi: 10.1016/j.heliyon.2024.e40729.
53. Gu D, Xia Y, Ding Z, et al. Inflammation in the peripheral nervous system after injury. *Biomedicines* 2024;12:1–15. doi: 10.3390/biomedicines12061256.

54. Sharifi M, Kamalabadi-Farahani M, Salehi M, Ebrahimi-Brough S, Alizadeh M. Recent perspectives on the synergy of mesenchymal stem cells with micro/nano strategies in peripheral nerve regeneration—a review. *Front Bioeng Biotechnol* 2024;12:1–21. doi: 10.3389/fbioe.2024.1401512.
55. Mushtaq M, Zineldeen DH, Mateen MA, Haider KH. mesenchymal stem cells’ “garbage bags” at work: treating radial nerve injury with mesenchymal stem cell-derived exosomes. *World J Stem Cells* 2024;16:467–78. doi: 10.4252/wjsc.v16.i5.467.
56. Li Q, Zhang F, Fu X, Han N. Therapeutic potential of mesenchymal stem cell-derived exosomes as nanomedicine for peripheral nerve injury. *Int J Mol Sci* 2024;25:7882. doi: 10.3390/ijms25147882.
57. Felici N, Alban A. Timing of surgery in peripheral nerve injury of the upper extremity. *J Hand Surg Eur* 2024;49:712–20. doi: 10.1177/17531934241240867.
58. Widodo W, Aprilya D, Satria O. Regenerative medicine: a new horizon in peripheral nerve injury and repair. *Orthop Rev (Pavia)* 2025;17:1–9. doi: 10.52965/001c.133572.
59. Aldali F, Deng C, Nie M, Chen H. Advances in therapies using mesenchymal stem cells and their exosomes for treatment of peripheral nerve injury: state of the art and future perspectives. *Neural Regen Res* 2025;20:3151–71. doi: 10.4103/NRR.NRR-D-24-00235.
60. Zhou H, He Y, Xiong W, et al. MSC based gene delivery methods and strategies improve the therapeutic efficacy of neurological diseases. *Bioact Mater* 2023;23:409–37. doi: 10.1016/j.bioactmat.2022.11.007.
61. Hammam IA, Winters R, Hong Z. Advancements in the application of biomaterials in neural tissue engineering: a review. *Biomed Eng Adv* 2024;8:100132. doi: 10.1016/j.bea.2024.100132.
62. Iwai T, Ikeguchi R, Aoyama T, et al. Nerve regeneration using a Bio 3D conduit derived from umbilical cord-derived mesenchymal stem cells in a rat sciatic nerve defect model. *PLoS One* 2024;19:1–17. doi: 10.1371/journal.pone.0310711.
63. Moghassemi S, Nikanfar S, Dadashzadeh A, et al. The revolutionary role of placental derivatives in biomedical research. *Bioact Mater* 2025;49:456–85. doi: 10.1016/j.bioactmat.2025.03.011.
64. Da Silva K, Kumar P, Choonara YE. The paradigm of stem cell secretome in tissue repair and regeneration: present and future perspectives. *Wound Repair Regen* 2025;33:1–32. doi: 10.1111/wrr.13251.
65. Namini MS, Daneshimehr F, Beheshtizadeh N, et al. Cell-free therapy based on extracellular vesicles: a promising therapeutic strategy for peripheral nerve injury. *Stem Cell Res Ther* 2023;14:1–18. doi: 10.1186/s13287-023-03467-5.
66. Nevado-Sánchez E, Rodríguez-Díaz M, Núñez-Rodríguez S, et al. Effectiveness of stem cell secretomes in the regeneration and functional recovery of severed nerves in patients with nerve injuries: a systematic review. *Cells* 2025;14:1–18. doi: 10.3390/cells14070492.
67. Wu S, Sun S, Fu W, Yang Z, Yao H, Zhang Z. The role and prospects of mesenchymal stem cells in skin repair and regeneration. *Biomedicines* 2024;12:743. doi: 10.3390/biomedicines12040743.
68. Han Y, Yang J, Fang J, et al. The secretion profile of mesenchymal stem cells and potential applications in treating human diseases. *Signal Transduct Target Ther* 2022;7:1–19. doi: 10.1038/s41392-022-00932-0.
69. Kumar MA, Baba SK, Sadida HQ, et al. Extracellular vesicles as tools and targets in therapy for diseases. *Signal Transduct Target Ther* 2024;9. doi: 10.1038/s41392-024-01735-1.
70. Bhol NK, Bhanjadeo MM, Singh AK, et al. The interplay between cytokines, inflammation, and antioxidants: mechanistic insights and therapeutic potentials of various antioxidants and anti-cytokine compounds. *Biomed Pharmacother* 2024;178:117177. doi: 10.1016/j.biopha.2024.117177.
71. Everts PA, Lana JF, Onishi K, et al. Angiogenesis and tissue repair depend on platelet dosing and bioformulation strategies following orthobiological platelet-rich plasma procedures: a narrative review. *Biomedicines* 2023;11:1922. doi: 10.3390/biomedicines11071922.
72. Wechsler ME, Rao V V., Borelli AN, Anseth KS. Engineering the msc secretome: a hydrogel focused approach. *Adv Health Mater* 2021;10:1–17. doi: 10.1002/adhm.202001948.
73. Pacilio S, Lombardi S, Costa R, et al. Role of perinatal stem cell secretome as potential therapy for muscular dystrophies. *Biomedicines* 2025;13:1–17. doi: 10.3390/biomedicines13020458.
74. Miron RJ, Estrin NE, Sculean A, Zhang Y. Understanding exosomes: part 2—emerging leaders in regenerative medicine. *Periodontol 2000* 2024;94:257–414. doi: 10.1111/prd.12561.
75. Wu SH, Zhou ZS, Li Y, Jiang J. Advancements in diabetic foot ulcer research: focus on mesenchymal stem cells and their exosomes. *Heliyon* 2024;10:e37031. doi: 10.1016/j.heliyon.2024.e37031.
76. Bina V, Brancato AM, Caliozna L, et al. Mesenchymal stem cells and secretome as a new possible approach to treat cartilage damage: an in vitro study. *Biomolecules* 2024;14:1068. doi: 10.3390/biom14091068.

77. Yang X, Tian S, Fan L, et al. Integrated regulation of chondrogenic differentiation in mesenchymal stem cells and differentiation of cancer cells. *Cancer Cell Int* 2022;22:1–13. doi: 10.1186/s12935-022-02598-8.
78. Yari H, Mikhailova M V, Mardasi M, et al. Emerging role of mesenchymal stromal cells (MSCs)-derived exosome in neurodegeneration-associated conditions: a groundbreaking cell-free approach. *Stem Cell Res Ther* 2022;13:1–23. doi: 10.1186/s13287-022-03122-5.
79. Izhiman Y, Esfandiari L. Emerging role of extracellular vesicles and exogenous stimuli in molecular mechanisms of peripheral nerve regeneration. *Front Cell Neurosci* 2024;18:1–21. doi: 10.3389/fncel.2024.1368630.
80. Lyamina S, Baranovskii D, Kozhevnikova E, et al. mesenchymal stromal cells as a driver of inflammaging. *Int J Mol Sci* 2023;24:6372. doi: 10.3390/ijms24076372.
81. Beheshtizadeh N, Gharibshahian M, Bayati M, et al. Vascular endothelial growth factor (VEGF) delivery approaches in regenerative medicine. *Biomed Pharmacother* 2023;166:115301. doi: 10.1016/j.biopha.2023.115301.
82. Huang Y, Wu L, Zhao Y, et al. Schwann cell promotes macrophage recruitment through il-17b/il-17rb pathway in injured peripheral nerves. *Cell Rep* 2024;43:113753. doi: 10.1016/j.celrep.2024.113753.
83. Torizal FG, Kerans FFA, Khumaira A. Production of mesenchymal stem cell derived-secretome as cell-free regenerative therapy and immunomodulation: a biomufacturing perspective. *Biocell.* 2022;46:1885–91. doi: 10.32604/biocell.2022.019591.
84. Yamaguchi N, Horio E, Sonoda J, et al. Immortalization of mesenchymal stem cells for application in regenerative medicine and their potential risks of tumorigenesis. *Int J Mol Sci* 2024;25:1–26. doi: 10.3390/ijms252413562.
85. Chouaib B, Haack-Sørensen M, Chaubron F, Cuisinier F, Collart-Dutilleul PY. Towards the standardization of mesenchymal stem cell secretome-derived product manufacturing for tissue regeneration. *Int J Mol Sci* 2023;24:12594. doi: 10.3390/ijms241612594.
86. Lu P, Ruan D, Huang M, et al. Harnessing the potential of hydrogels for advanced therapeutic applications: current achievements and future directions. *Signal Transduct Target Ther* 2024;9:166. doi: 10.1038/s41392-024-01852-x.
87. Kaur H, Gogoi B, Sharma I, et al. Hydrogels as a potential biomaterial for multimodal therapeutic applications. *Mol Pharm* 2024;21:4827–48. doi: 10.1021/acs.molpharmaceut.4c00595.
88. Chen H, Xu J, Sun J, et al. Recent advances on thermosensitive hydrogels-mediated precision therapy. *Asian J Pharm Sci* 2024;19:100911. doi: 10.1016/j.ajps.2024.100911.
89. Pablos JL, Lozano D, Manzano M, Vallet-Regí M. Regenerative medicine: hydrogels and mesoporous silica nanoparticles. *Mater Today Bio* 2024;29:101342. doi: 10.1016/j.mtbio.2024.101342.
90. Li F, Zhang J, Yi K, et al. Delivery of stem cell secretome for therapeutic applications. *ACS Appl Bio Mater* 2022;5:2009–30. doi: 10.1021/acsabm.1c01312.
91. Raspa A, Gelain F. mimicking extracellular matrix via engineered nanostructured biomaterials for neural repair. *Curr Neuropharmacol* 2020;19:2110–24. doi: 10.2174/1570159X18666201111111102.
92. Liang J, Liu P, Yang X, et al. Biomaterial-based scaffolds in promotion of cartilage regeneration: recent advances and emerging applications. *J Orthop Translat* 2023;41:54–62. doi: 10.1016/j.jot.2023.08.006.
93. Krishani M, Shin WY, Suhaimi H, Sambudi NS. Development of scaffolds from bio-based natural materials for tissue regeneration applications: a review. *Gels* 2023;9:100. doi: 10.3390/gels9020100.
94. Aoki K, Ideta H, Komatsu Y, et al. Bone-regeneration therapy using biodegradable scaffolds: calcium phosphate bioceramics and biodegradable polymers. *Bioengineering* 2024;11:180. doi: 10.3390/bioengineering11020180.
95. Wang M, Xu Y, Cao L, et al. Mechanical and biological properties of 3d printed bone tissue engineering scaffolds. *Front Bioeng Biotechnol* 2025;13:1–23. doi: 10.3389/fbioe.2025.1545693.
96. Baptista CJM, Rocha SCM. Biochemical properties of amniotic membrane. In: Mamede AC, Carvalho MJ, Abrantes A, Laranjo M, Maia C, Botelho MF, editors. *Amniotic membrane : origin, characterization, and medical application*. Dordrecht: Springer Science+Business Media; 2015; pp. 19–40. doi: 10.1007/978-94-017-9975-1\_2.
97. Zhou T, Yuan Z, Weng J, et al. Challenges and advances in clinical applications of mesenchymal stromal cells. *J Hematol Oncol* 2021;14:1–24. doi: 10.1186/s13045-021-01037-x.
98. Gerdfaramarzi MS, Bazmi S, Kiani M, Afshar L, Fadavi M, Enjoo SA. Ethical challenges of cord blood banks: a scoping review. *J Med Life* 2022;15:735–41. doi: 10.25122/jml-2021-0162.
99. Rebelatto CLK, Boldrini-Leite LM, Daga DR, et al. Quality control optimization for minimizing security risks associated with mesenchymal stromal cell-based product development. *Int J Mol Sci* 2023;24: 12955. doi: 10.3390/ijms241612955.

100. Capelli C, Cuofano C, Pavoni C, et al. Potency assays and biomarkers for cell-based advanced therapy medicinal products. *Front Immunol* 2023;14:1–18. doi: 10.3389/fimmu.2023.1186224.
101. Partan RU, Putra KM, Kusuma NF, et al. Umbilical cord mesenchymal stem cell secretome improves clinical outcomes and changes biomarkers in knee osteoarthritis. *J Clin Med* 2023;12:7138. doi: 10.3390/jcm12227138.
102. Lubis AMT, Aprianto P, Pawitan JA, Priosoeryanto BP, Dewi TIT, Kamal AF. Intra-articular injection of secretome, derived from umbilical cord mesenchymal stem cell, enhances the regeneration process of cartilage in early-stage osteo-arthritis: an animal study. *Acta Orthop* 2023;94:300-6. doi: 10.2340/17453674.2023.12359.
103. Li S, Rong Q, Zhou Y, Che Y, Ye Z, Liu J, Wang J, Zhou M. Osteogenically committed hUCMSCs-derived exosomes promote the recovery of critical-sized bone defects with enhanced osteogenic properties. *APL Bioeng* 2024;8:016107. doi: 10.1063/5.0159740.
104. Jia S, Yang T, Gao S, Bai L, et al. Exosomes from umbilical cord mesenchymal stem cells ameliorate intervertebral disc degeneration via repairing mitochondrial dysfunction. *J Orthop Translat* 2024;46:103-15. doi: 10.1016/j.jot.2023.10.004.



This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License

---